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REVIEW

Prostaglandins and Leukotrienes: Advances in Eicosanoid Biology

Colin D. Funk

Prostaglandins and leukotrienes are potent eicosanoid lipid mediators derived from phospholipase-released arachidonic acid that are involved in numerous homeostatic biological functions and inflammation. They are generated by cyclooxygenase isozymes and 5-lipoxygenase, respectively, and their biosynthesis and actions are blocked by clinically relevant nonsteroidal anti-inflammatory drugs, the newer generation coxibs (selective inhibitors of cyclooxygenase-2), and leukotriene modifiers. The prime mode of prostaglandin and leukotriene action is through specific G protein–coupled receptors, many of which have been cloned recently, thus enabling specific receptor agonist and antagonist development. Important insights into the mechanisms of inflammatory responses, pain, and fever have been gleaned from our current understanding of eicosanoid biology.

A discovery chain culminating in one of the most important classes of lipid mediators, known as eicosanoids (from the Greek eicosa = twenty; for twenty carbon fatty acid derivatives), was initiated in 1930 with two seminal, though seemingly unrelated, laboratory observations (1-3). The first of these found that exclusion of fat from the diet of rats led to growth retardation, reproductive disturbances, scaly skin, kidney lesions, and excessive water consumption, which led to the discovery of essential fatty acids. The second identified a factor with fatty acid properties and vasodepressor and smooth muscle-stimulating activity that was termed "prostaglandin." Bergström and Samuelsson, some 30 years later, linked these observations when they elucidated the structures of the "classical" prostaglandins and demonstrated that they were produced from an essential fatty acid, arachidonic acid (4). Thus began an era of eicosanoid research.

The diverse and potent biological actions of prostaglandins on almost all organs stimulated research on these fascinating molecules over the ensuing four decades. In 1971, Vane discovered that aspirin-like drugs, known for their analgesic, antipyretic, and anti-inflammatory actions, could inhibit prostaglandin biosynthesis (5). Soon thereafter, the platelet proaggregatory and vasoconstrictor molecule thromboxane A2 was elucidated, followed by the isolation of vascular wall-synthesized prostacyclin, which counteracts thromboxane action. Prostaglandins were found to induce labor or act as abortifacients. From 1975 to 1980, the leukotriene biosynthetic pathway was compiled, which demonstrated that leukotrienes are potent lipid mediators associated with asthma and allergic reactions (6). Lastly, Nobel Prizes related to

the eicosanoid field were awarded (7).

The composition of the eicosanoid family has expanded immensely in the past two decades to include virtually all long-chain oxygenated polyunsaturated products, whether formed enzymatically or nonenzymatically. Arachidonic acid ($20:4\omega 6$) is the premier eicosanoid precursor in mammalian cells. The quintessential properties of "true" eicosanoids are their stereochemical precision in formation and recognition, their potency in the nanomolar range in vitro, and their bona fide biological activities. This review will cover our current knowledge of two eicosanoid members, the prostaglandins and leukotrienes.

Biosynthesis of Prostaglandins

Prostaglandins are formed by most cells in our bodies and act as autocrine and paracrine lipid mediators (i.e., they signal at or immediately adjacent to their site of synthesis). They are not stored but are synthesized de novo from membrane-released arachidonic acid when cells are activated by mechanical trauma or by specific cytokine, growth factor, and other stimuli [e.g., collagen and adenosine diphosphate (ADP) in platelets, bradykinin and thrombin in endothelium]. A host of enzymes exquisitely regulate cellular levels of arachidonic acid, keeping it esterified until mobilized by phospholipases (PLA_2) . The control of arachidonic acid release from membranes has undergone several paradigm shifts in recent years with the continuing identification of new PLA₂ members (8). Despite this, type IV cytosolic PLA₂ (cPLA₂) remains the key player for eicosanoid production because cells lacking cPLA₂ are generally devoid of eicosanoid synthesis. Cell-specific and agonist-dependent events coordinate translocation of cPLA₂ to the nuclear envelope, endoplasmic reticulum (ER), and Golgi apparatus (9).

At the ER and nuclear membrane, arachidonic acid released by $cPLA_2$ is presented to prostaglandin H synthase (PGHS; referred to colloquially as COX for cyclooxygenase) and is then metabolized to an intermediate prostaglandin PGH₂ (Fig. 1). PGHS exists as two isoforms referred to as PGHS-1 (COX-1) and PGHS-2 (COX-2) (10). In simplistic terms, COX-1 is the enzyme responsible for basal, constitutive prostaglandin synthesis, whereas COX-2 is important in various inflammatory and "induced" settings. There are notable exceptions to this oversimplification, but in general this classification has aided the rapid advancement in this field since the discovery of COX-2 10 years ago. The COX enzymes are monotopically inserted in the ER and nuclear membrane with the substratebinding pocket precisely orientated to take up released arachidonic acid. The crystal structures of COX-1 and COX-2 are remarkably similar, with one notable amino acid difference that leads to a larger "side-pocket" for substrate access in COX-2 (10).

The coupling of PGH₂ synthesis to metabolism by downstream enzymes is intricately orchestrated in a cell-specific fashion. Thromboxane synthase is found in platelets and macrophages, prostacyclin synthase is found in endothelial cells and PGF synthase in uterus, and two types of PGD synthase are found in brain and mast cells. Microsomal PGE synthase (mPGES), a member of the MAPEG (membrane-associated proteins in eicosanoid and glutathione metabolism) family, is responsible for PGE₂ synthesis (11). Coordinate induction of multiple enzymes in the prostanoid pathway, in particular mPGES and COX-2, in inflammatory settings is a current concept being developed (12).

Biosynthesis of Leukotrienes

In contrast to prostaglandins, leukotrienes are made predominantly by inflammatory cells like polymorphonuclear leukocytes, macrophages, and mast cells. Cellular activation by immune complexes, bacterial peptides, and other stimuli elicit a sequence of events that include cPLA₂ and 5-lipoxygenase (5-LO) translocations to the nuclear envelope (Fig. 2). 5-LO, a nonheme iron dioxygenase, is the key enzyme in this cascade and is located in the nucleus in some cell types and the cytosol of others (13). 5-LO possesses a NH₂-terminal domain that binds two calcium ions, similar to the β -sandwich C2 domains of cPLA, and protein kinase C, and a large catalytic domain that binds iron (14, 15). It transforms released arachidonic acid to the epoxide LTA₄ with the concerted efforts of 5-lipoxygenase-activating protein (FLAP). There are questions as to how this cascade of events takes

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place. The directed 5-LO translocation process could be governed by specific motifs in its structure (e.g., SH3 binding region) through protein-protein or protein-cytoskeleton interactions or by the intrinsic predilection of its NH₂-terminal domain to seek out phosphatidylcholine-rich lipid domains. FLAP may retrieve the substrate and transfer it to 5-LO, but the mechanisms remain obscure.

LTA4 undergoes transformation by one or more of three possible fates depending on the cellular context: hydrolysis, conjugation with glutathione, or transcellular metabolism to generate bioactive eicosanoids (16). Hydrolytic attack of LTA₄ by leukotriene A₄ hydrolase (LTA4H) in the cytoplasm, and potentially in the nucleus, yields LTB₄, a potent neutrophil chemoattractant and stimulator of leukocyte adhesion to endothelial cells (6, 13). LTA₄H is a bifunctional zinc-containing enzyme with epoxide hydrolase and aminopeptidase activities. The three-domain crystal structure of LTA₄H, with a catalytic domain highly related to thermolysin and a COOH-terminal domain with armadillo-like repeats (17), points to an interesting evolutionary heritage. Because LTA₄H is found in yeast (18), predating the appearance of other leukotriene biosynthetic enzymes, it may have had, at one time, exclusively aminopepti-

Fig. 1. Prostaglandin synthesis and actions. A "generic" cell when activated by mechanical trauma, cytokines, growth factors, or various inflammatory stimuli triggers signaling, including type IV cytosolic phospholipase (cPLA₂) translocation to ER and nuclear membranes, arachidonic acid release from membrane lipids and metabolism by COX-1 or COX-2 to the intermediate PGH₂. Other PLA₂ subtypes could be involved in arachidonic acid release for eicosanoid synthesis but are not shown here. De novo COX-2 enzyme synthesis can be induced by a host of factors (top) to reinforce prostaglandin (PG) formation. In a cell-type restricted fashion, a heterogeneous family of PGH metabolizing enzymes can form $PGE_{2'}^{2}$ $PGD_{2'}$ $PGF_{2\alpha'}$ PGI_{2} (prostacyclin) and TxA, (thromboxane). These prostaglandins may undergo facilitated transport from the cell through a known prostaglandin transporter (PGT) or other carrier to exert autocrine or paracrine actions on a family of prostaglandin receptors EP₁, EP₂, EP₃, EP₄ DP_1 , DP_2 , FP, IP, TP_{α} , and TP_{β} on the cell types indicated. Only a few of the many diverse activities of prostaglandins are shown here. Prostaglandins could potentially enter the nucleus and activate nuclear hormone receptors such as PPAR-γ. PGES, PGE synthase; PGDS, PGD synthase; PGFS, PGF synthase; PGIS, prostacyclin synthase; TxS, thromboxane synthase. VSMC is vascular smooth muscle cell. OVLT in POA is

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dase or other noneicosanoid related functions.

 LTA_4 conjugation with glutathione to form LTC_4 at the nuclear envelope is catalyzed by LTC_4 synthase (LTC_4 H) (19), another member of the MAPEG family. LTC_4 is transported out of the cell by transporters such as the multidrug resistance–associated protein (MRP1), a process that apparently regulates dendritic cell migration to lymph nodes (20). The peptide moiety of LTC_4 is subjected to extracellular metabolism, forming LTD_4 and LTE_4 . These three leukotrienes comprise the cysteinyl leukotrienes, or an entity described more than 60 years ago as "slow-reacting substance of anaphylaxis" for its slow and sustained smooth muscle contracting abilities (6).

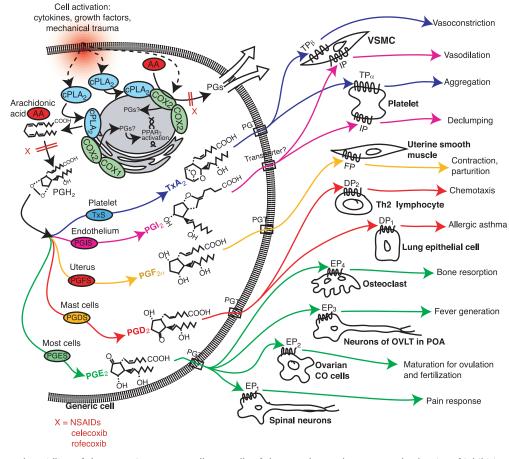
Mechanisms of Prostaglandin Action

Prostaglandins are released from cells predominantly by facilitated transport through a known prostaglandin transporter (PGT) of the organic anion transporter polypeptide family, and potentially by other uncharacterized transporters (21). Due to the evanescent nature of thromboxane and prostacyclin (which have half-lives on the order of seconds to a few minutes), these compounds must act near their sites of synthesis. There are at least 9 known prostaglandin receptor forms in mouse and man, as well as several

additional splice variants with divergent carboxy termini (22). Four of the receptor subtypes bind PGE₂ (EP₁-EP₄), two bind PGD₂ (DP₁ and DP_2) (23–25), and the receptors that bind PGF₂₀, PGI₂, and TxA₂ (FP, IP, and TP, respectively) each derive from a single gene. The prostaglandin receptors belong to three clusters (on the basis of homology and signaling attributes rather than by ligand-binding properties) within a distinct subfamily of the G proteincoupled receptor (GPCR) superfamily of seventransmembrane spanning proteins. The lone exception is DP₂, which is a member of the chemoattractant receptor subgrouping. The "relaxant" receptors IP, DP1, EP2, and EP4 form one cluster, signaling through G_s-mediated increases in intracellular cyclic adenosine monophosphate (cAMP); the "contractile" receptors EP1, FP, and TP form a second group that signals through G_a-mediated increases in intracellular calcium. The EP₃ receptor is regarded as an "inhibitory" receptor that couples to G_i and decreases cAMP formation. Although most of the prostaglandin GPCRs are localized at the plasma membrane, some are situated at the nuclear envelope (26).

Mechanisms of Leukotriene Action

Leukotrienes also act at distinct GPCRs, four of which have been characterized to date (27-30)



the organum vasculosa lamina terminalis at the midline of the preoptic area. CO cells are cells of the cumulus oophorus. X marks the site of inhibition by NSAIDs (aspirin, ibuprofen, indomethacin) and the coxibs celecoxib (Celebrex) and rofecoxib (Vioxx).

(Fig. 2). The high-affinity B-LT₁ receptor on leukocytes binds LTB_4 in the subnanomolar range and elicits a pertussis toxin-sensitive Gilinked chemotactic response. High concentrations of LTB4 and signaling through Ga coupling stimulates neutrophil secretion. A recently characterized B-LT₂ receptor that binds LTB₄ with much lower affinity than B-LT₁ displays a widespread tissue distribution pattern, and its function is presently unknown. Interestingly, the genes for B-LT1 and B-LT2 reside intertwined, one within the promoter of the other (28). Two subtypes of cysteinyl leukotriene receptors, CysLT₁ and CysLT₂, mediate the actions of LTC4 and LTD4. CysLT1 is found on airway smooth muscle cells (29) and vascular endothelial cells (31) promoting bronchoconstriction and up-regulation of cell adhesion molecules, respectively. CysLT1 may also masquerade as a pyrimidinergic (UDP) receptor but the significance of this finding remains to be determined (32). CysLT₂, originally found in pulmonary vein preparations by pharmacological assays, is detected in spleen, Purkinje fibers of the heart, and discrete regions of the adrenal gland by molecular methods. Leukotriene functions in these tissues are unknown, so fruitful avenues for future research will arise.

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GPCR, PPARs, or Both?

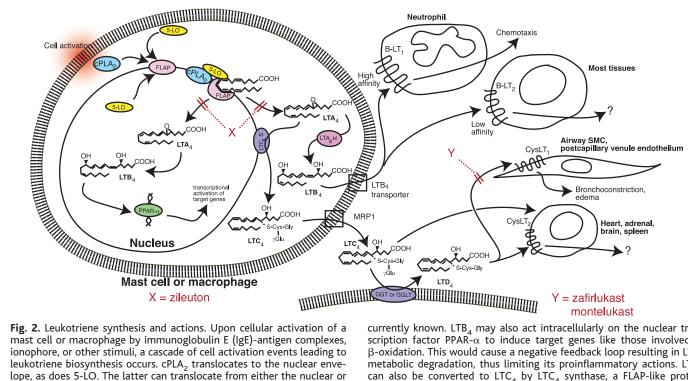
Do prostaglandins and leukotrienes exert their actions solely through GPCR? Peroxisomal proliferator-activated receptors (PPARs) can bind and be activated by a variety of eicosanoids; PPAR- α by LTB₄ and 8(S)-HETE, PPAR- γ by 15-deoxy-delta-12,14-PGJ₂ (a dehydration metabolite of PGD_2), and PPAR- δ by prostacyclin analogs (33-37). However, the approach for ligand discovery was not based on known eicosanoid biochemistry. Problematic issues relate to cell-specific eicosanoid biosynthetic patterns, whether certain ligands are formed in vivo (e.g., 15-deoxy-delta-12,14-PGJ₂), and the concentrations of ligand needed to activate responses (micromolar range versus nanomolar for conventional eicosanoid GPCR) (22). Whether eicosanoids are bona fide endogenous PPAR ligands has yet to be resolved with rigorous analytical methods and testing of COX, 5-LO, and other eicosanoid-deficient mice.

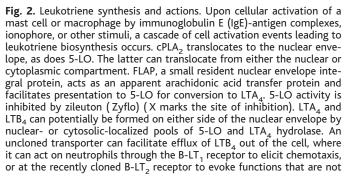
Drugs Affecting Prostaglandin and Leukotriene Formation and Action

Nonsteroidal anti-inflammatory drugs (NSAIDs; e.g., aspirin, indomethacin, ibuprofen), known to block PGHS-derived prosta-

glandin synthesis, are firmly entrenched in the common man's armamentarium of analgesics and anti-inflammatories. Although the mechanism of COX inhibition by NSAIDs is rarely disputed, some NSAIDs affect the transcription factors nuclear factor kappa B (NF-KB) and PPAR family members; however, higher concentrations are required than those that effectively block COX activity (38, 39). Aspirin remains the sole member of this class of drugs with a unique mechanism of action on COX by covalently acetylating a serine residue. This blocks proper substrate access and orientation at the active site. The coxibs (selective inhibitors of COX-2), celecoxib (Celebrex) and rofecoxib (Vioxx), are newer COX-2 specific drugs that have been used clinically for the past 2 years in arthritis and other pain symptom management (40). The second-generation coxibs, valdecoxib and etoricoxib, are undergoing clinical development.

Leukotriene modifiers or antileukotrienes constitute 5-lipoxygenase inhibitors [zileuton (Zyflo)] and CysLT₁ receptor antagonists [zafirlukast (Accolate) and montelukast (Singulair)] used clinically in long-term maintenance of asthma control (41). Are leukotriene modifiers an important asthma therapy? There is much debate about their clinical efficacy (42). There





currently known. LTB₄ may also act intracellularly on the nuclear transcription factor PPAR- α to induce target genes like those involved in β -oxidation. This would cause a negative feedback loop resulting in LTB₄ metabolic degradation, thus limiting its proinflammatory actions. LTA₄ can also be converted to LTC₄ by LTC₄ synthase, a FLAP-like protein found in the nuclear envelope. The multidrug resistance-associated protein (MRP1) can facilitate transfer of LTC₄ out of the cell, where it is metabolized by extracellular-localized γ -glutamyl transpeptidase (GGT) or γ -glutamyl leukotrienase (GGLT) to LTD₄. On airway smooth muscle cells (SMC) and postcapillary venule endothelial cells, LTD₄ can activate CysLT₁ receptors to cause bronchoconstriction and edema. The clinically important drugs in asthma, montelukast (Singulair) and zafirlukast (Accolate), block this binding step (Y marks the site of inhibition). LTC₄ or LTD₄ may also bind CysLT₂ receptors found in a variety of tissues.

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is well-documented effectiveness in exerciseinduced asthma and aspirin-intolerant asthma. Clinical trials show bronchodilatory effects beyond those provided by β agonists, as well as reduced eosinophil numbers in sputum. But there are also definite nonresponder patients, which might be explained by nonleukotriene dependent asthma mechanisms or by pharmacogenetic factors. In fact, the therapeutic response to a 5-LO inhibitor can vary with 5-LO promoter polymorphisms containing variable numbers of GC-boxes capable of binding Sp1 and Egr-1 transcription factors (43, 44). Leukotriene modifiers provide a steroid-sparing benefit in mild to moderate asthmatics, although questions remain regarding the additional advantage of adding an antileukotriene to traditional therapies (B agonists, corticosteroids, theophyllines) in chronic persistant asthma (42, 45, 46).

The Multifaceted Roles of Eicosanoids

The renewed growth and rapid advance in the eicosanoid field over recent years is attributed in part to studies carried out with knockout mice (22, 47, 48). Virtually all genes encoding enzymes and receptors in the prostaglandin and leukotriene pathways have been disrupted by gene targeting. The studies are unraveling novel eicosanoid actions, confirming long-held viewpoints, and providing provocative data for further in-depth research. Eicosanoids are implicated in functions in practically every organ, tissue, and cell in our bodies (see some examples in Figs. 1 and 2). One prominent area of interest is the role of eicosanoids in pain, fever, and inflammation.

Inflammation. Scientists have been grappling for years over the specific mechanisms of how prostaglandins mediate their effects on the cardinal signs of acute inflammation: pain, vasodilation (swelling and redness), and fever. COX-1 is expressed in nearly all tissues, whereas COX-2 is absent in most (some exceptions are the glomerulus and certain brain regions) until induced by various inflammatory insults in monocytes or mast cells or by shear stress in endothelium. In most instances, COX-1 expression is marginally affected by inflammatory stimuli. However, exceptions to the "constitutive" mode of COX-1 prostanoid synthesis are known (e.g., both COX-1 and COX-2 are expressed in the inflamed synovia of joints). Most of the traditional NSAIDs do not distinguish between the two COX isoforms. Coxibs, however, were developed specifically with the promise that they would selectively block synthesis of "proinflammatory" prostaglandins derived from the induced COX-2 enzyme while leaving intact the COX-1-derived "homeostatic" prostaglandins involved in renal water and electrolyte balance, gastric cytoprotection, and platelet aggregation (40). Two years of clinical use in pain management indicate that coxibs are as effective as traditional nonselective NSAIDs

and also reveal a 50 % reduction in adverse gastrointestinal events (40). Although indications of potentially deleterious actions of COX-2 inhibitors (e.g., causing acute tubulointerstitial nephritis or decreased cardioprotection) have been reported (49, 50), the case to support an increased incidence of adverse events compared with traditional NSAIDs has not been developed.

Vasodilation and increased permeability of postcapillary venules, early events in the inflammatory response, reflect the effects of COX-2-derived prostaglandins and leukotrienes at sites of inflammation. Prostaglandins synergize with other mediators (e.g., bradykinin, histamine) to elicit enhanced vascular permeability and edema. These molecules can be viewed within the context of a complex milieu of parenchymal and inflammatory cells, an array of cytokine and other noneicosanoid mediators, and extracellular matrix interactions combined with the overall physiological status of the host. To complicate matters, prostaglandins may act as both proinflammatory and anti-inflammatory mediators depending on the context, which is due in part to the array of EP-type prostaglandin receptors with opposing signal transduction pathways. How tissues and cells sort out the mixed signals has been reviewed recently (48). The temporal sequence of events in acute inflammation may be governed by eicosanoid profile switching such that eicosanoids made during the initial phase are gradually replaced by other lipid mediators in the resolution phase (51). In vitro evidence indicates that monocytes and/or macrophages can undergo a shift in eicosanoid products (52, 53), a process perhaps mediated by altered gene expression of the synthases downstream of COX-1 and COX-2 or by specific compartmentalization of the enzymes to various stimuli. Combined data from several murine inflammation models support a complex regulatory network in eicosanoid signaling (48, 51, 54).

The 5-LO pathway leading to leukotriene formation has long been recognized as a proinflammatory cascade. LTB₄ promotes neutrophil chemotaxis and adhesion to vascular endothelium through specific integrins. The cysteinyl leukotrienes cause plasma leakage from postcapillary venules and enhance mucus secretion. LTD₄ and another 5-LO-derived eicosanoid, 5-oxo-ETE, are eosinophil chemoattractants. The use of 5-lipoxygenase, FLAP-, LTA₄H-, and LTC₄S-deficient mice has enabled a detailed examination of the leukotrienes in murine models, firmly establishing their roles in allergic inflammation.

Pain. The antinociceptive effects of NSAIDs are well recognized because prostaglandins elicit a hyperalgesic response, or increased sensitivity, to touch by sensitiz-

ing the free end of pain neurons in peripheral inflammation. Prostaglandins act both at peripheral sensory neurons and at central sites within the spinal cord and brain to evoke hyperalgesia. Recent data support involvement of both PGI₂ by IP and PGE₂ by EP₁ receptors in pain (22, 48, 55). Prostaglandins are also involved in allodynia, a pain response to a usually nonpainful stimulus. Circulating interleukin-1ß cytokine, which originates at a peripheral injury site and cannot pass the blood-brain barrier, induces both COX-2 and mPGES activities in cells lining the barrier (56, 57). PGE, then enters the brain and cerebrospinal fluid and induces prostanoid receptor activation on neurons and microglia. This increases neuronal excitability and leads to nonpainful stimuli becoming painful, basically converting a peripheral injury to a central pain response without nerve impulse transmission. Specific EP1 or IP receptor antagonists could be useful in pain management. Whether they offer any benefit over coxibs or NSAIDs remains to be discovered.

Fever. The potent antipyrogenic effects of NSAIDs have provided strong evidence for a role of prostaglandins in fever, but the mechanisms have remained obscure until recently (55, 58). Bacterial lipopolysaccharides and other marauding challenges induce cytokine networks that cause fever. Subsequently, these stimulate the neural pathways that raise body temperature. In response to both exogenous and endogenous pyrogens, cytokinereleased PGE, derived from COX-2 in the organum vasculosa lamina terminalis (OVLT), at the midline of the preoptic area, mediates the febrile response. This occurs through the EP3 receptor expressed in neurons surrounding the OVLT, a region exquisitely sensitive to exogenous PGE2-induced pyrexia.

Prospects

Where is the eicosanoid field heading in the next few years? Interest in the area tends to undergo cyclical periods of grandeur and demise. A strong resurgent period is upon us, after the introduction of COX-2 inhibitors into the clinics, the cloning of four leukotriene receptor subtypes, and the characterization of prostaglandin receptor knockout mice. What lies on the horizon? Sorting out the events in the regulation of prostaglandin and leukotriene biosynthesis is definitely a priority. Are their specific microdomains to which cPLA₂ and 5-LO target? The compartmentalization of the enzymes within the eicosanoid pathways has received considerable attention but requires further refinement (10, 13). 5-LO undergoes dynamic movement in and out of the nucleus in a cell- and stimulus-dependent

manner. How does rapid, induced synthesis of COX-2 coordinate with cPLA₂, other secretory PLA₂s and downstream synthases during inflammation to produce a proinflammatory eicosanoid profile? Is there an orchestrated temporal change in lipid mediators during the resolution phase? Answers to these questions are forthcoming.

The development of specific agonists and antagonists for each of the prostaglandin and leukotriene receptors will provide important reagents for further defining the biological importance of this group of bioactive lipids. Reports on genetic variants of eicosanoid receptors and biosynthetic enzymes within the prostaglandin and leukotriene pathways have been scant. Elucidation of such variants and their potential relevance to inflammation or disease susceptibility and interindividual variations in drug response will be an area of active investigation.

Advances in eicosanoid biology certainly extend beyond the prostaglandins and leukotrienes. The hydroxy (HETE) (59), epoxy (EET) (60), and lipoxin eicosanoid molecules (61) are emerging areas as well. NSAIDs and coxibs may also turn out to be useful therapeutic agents in the treatment of Alzheimer's disease and certain cancers (62-64). Other lipoxygenase and cyclooxygenase products are implicated in atherogenesis (65, 66) and will also certainly receive attention in the years to come.

The eicosanoids, like no other set of lipid mediators, possess a vast array of biological actions in many different cell types. Eicosanoid molecules are truly a conundrum, paradoxically acting as both friend and foe. New insight into their roles in pain, inflammation, and disease and the development of novel therapeutics will undoubtedly arise in the near future.

References and Notes

- G. O. Burr, M. M. Burr, J. Biol. Chem. 86, 587 (1930).
 R. Kurzrok, C. C. Lieb, Proc. Soc. Exp. Biol. Med. 28, 268 (1930).
- 3. U. S. von Euler, J. Physiol. 81, 102 (1934).
- 4. S. Bergström, H. Danielsson, B. Samuelsson, *Biochim. Biophys. Acta* **90**, 207 (1964).
- 5. J. R. Vane, Nature New Biol. 231, 232 (1971).
- 6. B. Samuelsson, Science 220, 568 (1983).
- Samuelsson, Vane, and Bergström were awarded the prize in medicine or physiology in 1982 and E. J. Corey was awarded it in chemistry in 1990 (see www.nobel.se/medicine/laureates/1982/index.html and www.nobel.se/chemistry/laureates/1990/ index.html).
- D. A. Six, E. A. Dennis, *Biochim. Biophys. Acta* **1488**, 1 (2000).
- J. H. Evans, D. M. Spencer, A. Zweifach, C. C. Leslie, J. Biol. Chem. 276, 30150 (2001).
- W. L. Smith, D. L. DeWitt, R. M. Garavito, Annu. Rev. Biochem. 69, 145 (2000).
- 11. P. J. Jakobsson, S. Thorén, R. Morgenstern, B. Samuelsson, *Proc. Natl. Acad. Sci. U.S.A.* **96**, 7220 (1999).
- 12. J. A. Mancini et al., J. Biol. Chem. 276, 4469 (2001).
- 13. M. Peters-Golden, T. G. Brock, *FEBS Lett.* **487**, 323 (2001).
- 14. T. Hammarberg, P. Provost, B. Persson, O. Rådmark, J. Biol. Chem. 275, 38787 (2000).
- 15. X. S. Chen, C. D. Funk, *J. Biol. Chem.* **276**, 811 (2001). 16. K. Gronert, C. B. Clish, M. Romano, C. N. Serhan,
- Methods Mol. Biol. **120**, 119 (1999). 17. M. M. Thunnissen, P. Nordlund, J. Z. Haeggström,
- Nature Struct. Biol. 8, 131 (2001). 18. F. Kull, E. Ohlson, J. Z. Haeggström, J. Biol. Chem. 274, 34683 (1999).
- 19. J. F. Penrose, K. F. Austen, *Proc. Assoc. Am. Physicians* 111, 537 (1999).
- 20. D. F. Robbiani et al., Cell 103, 757 (2000).
- 21. V. L. Schuster, Annu. Rev. Physiol. 60, 221 (1998).
- S. Narumiya, G. A. FitzGerald, J. Clin. Invest. 108, 25 (2001).
- 23. H. Hirai et al., J. Exp. Med. 193, 255 (2001).
- 24. G. Monneret et al., Blood 98, 1942 (2001).
- 25. DP₁ and DP₂ designations have been used informally, but this nomenclature has not yet been approved. CRTH2 (chemoattractant receptor homologous molecule expressed on T helper 2 lymphocytes) is the original designation for DP₂.
- M. Bhattacharya et al., Proc. Natl. Acad. Sci. U.S.A., 95, 15792 (1998).
- 27. T. Yokomizo et al., Nature **387**, 620 (1997).
- 28. T. Yokomizo et al., J. Exp. Med. 192, 421 (2000).
- 29. K. R. Lynch et al., Nature **399**, 789 (1999).
- 30. C. E. Heise et al., J. Biol. Chem. 275, 30531 (2000).
- 31. K. Gronert et al., Am. J. Pathol. 158, 3 (2001).

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- E. A. Mellor, A. Maekawa, K. F. Austen, J. A. Boyce, Proc. Natl. Acad. Sci. U.S.A. 98, 7964 (2001).
- 33. K. Yu, et al., J. Biol. Chem. 270, 23975 (1995).
- 34. P. R. Devchand et al., Nature 384, 39 (1996).
- 35. S. A. Kliewer et al., Cell 83, 813 (1995)
- 36. B. M. Forman et al., Cell 83, 803 (1995).
- 37. R. A. Gupta et al., Proc. Natl. Acad. Sci. U.S.A. 97, 13275 (2000).
- T. C. He, T. A. Chan, B. Vogelstein, K. W. Kinzler, *Cell* 99, 335 (1999).
- B. Frantz, E. A. O'Neill, *Science* **270**, 2017 (1995).
 G. A. FitzGerald, C. Patrono, *N. Engl. J. Med.* **345**, 433
- (2001). 41. J. M. Drazen, E. Israel, P. M. O'Byrne. *N. Engl. J. Med.*
- **340**, 197 (1999). 42. R. A. Nathan, J. P. Kemp, *Ann. Allergy Asthma Immu-*
- nol. 86, 9 (2001). 43. C. D. Funk et al., Proc. Natl. Acad. Sci. U.S.A. 86,
- 2587 (1989). 44. J. R. Drazen *et al., Nature Genet.* **22**, 168 (1999).
- 45. D. S. Robinson, D. Campbell, P. J. Barnes, *Lancet* **357**, 2007 (2001).
- 46. R. H. Green, I. D. Pavord, Lancet 357, 1991 (2001).
- S. C. Austin, C. D. Funk, Prostaglandins Lipid Med. 58, 231 (1999).
- S. L. Tilley, T. M. Coffman, B. H. Koller, J. Clin. Invest. 108, 15 (2001).
- 49. J. L. Rocha, J. Fernandez-Alonzo, *Lancet* **357**, 1946 (2001).
- 50. M. Boers, Lancet **357**, 1222 (2001).
- 51. B. D. Levy et al., Nature Immunol. 2, 612 (2001).
- 52. P. S. Penglis *et al., J. Immunol.* **165**, 1605 (2000). 53. T. G. Brock, R. W. McNish, M. Peters-Golden, *J. Biol.*
- I. G. Brock, R. W. McNish, M. Peters-Golden, J. Biol Chem. 274, 11660 (1999).
- 54. D. W. Gilroy et al., Nature Med. 5, 698 (1999).
- Y. Sugimoto, S. Narumiya, A. Ichikawa, Prog. Lipid Res. 39, 289 (2000).
- 56. M. Ek et al., Nature 410, 430 (2001).
- 57. T. Bartfai, Nature **410**, 425 (2001).
- 58. F. Ushikubi et al., Nature 395, 281 (1998).
- V. A. Ziboh, C. C. Miller, Y. Cho, Prostaglandins Other Lipid Mediat. 63, 3 (2000).
- 60. D. C. Zeldin, J. Biol. Chem. 276, 36059 (2001).
- C. N. Serhan, E. Oliw, J. Clin. Invest. 107, 1481 (2001).
 A. Yermakova, M. K. O'Banion, Curr. Pharm. Des. 6,
- 1755 (2000). 63. K. E. Giercksky, Best Pract. Res. Clin. Gastroenterol.
- **15**, 821 (2001). 64. S. Wegger *et al.*, *Nature* **414**, 212 (2001).
- 65. C. D. Funk, T. Cyrus, Tr. Cardiovasc. Med. 11, 116
- (2001). 66. G. A. FitzGerald *et al., Ann. Med.* **32** (suppl. 1), 21
- (2000). 57. I regret being unable to cite all relevant references,
- Fregret being unable to cite all relevant references, due to space constraints. Supported by NIH grants HL58464, HL53558, and GM63130.
- Lysophospholipids—Receptor Revelations

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Upon cell activation, membrane phospholipids are metabolized into potent lysophospholipid (LP) mediators, such as sphingosine 1-phosphate and lysophosphatidic acid. LPs fulfill signaling roles in organisms as diverse as yeast and humans. The recent discovery of G protein–coupled receptors for LPs in higher eukaryotes, and their involvement in regulating diverse processes such as angiogenesis, cardiac development, neuronal survival, and immunity, has stimulated growing interest in these lipid mediators. LP receptor biology has generated insights into fundamental cellular mechanisms and may provide therapeutic targets for drug development.

Glycerol-based and sphingosine-based phospholipids are abundant structural components of cellular membranes; however, they are metabolized into polar metabolites such as eicosanoids and lysophospholipids (LPs) (1). The latter includes lysophosphatidic acid (LPA), lysophosphatidylcholine (LPC), sphingosylphosphoryl choline (SPC), and sphingosine 1-phosphate (S1P). ¹However, in contrast to the eicosanoids, whose critical roles in normal physiology and disease are underscored by the current widespread clini2007

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